

A photograph of a pond with several purple flowers and large green lily pads. The water is dark, and the scene is brightly lit. The text is overlaid on a semi-transparent green rectangular area in the center of the image.

Developing eDNA Tools For Native and Invasive Aquatic Plant Detection in Maine

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Aquatic plants are important to wildlife & water quality

- Protect water quality
- Stabilize shoreline and sediments
- Provide oxygen and play key role in nitrogen cycle
- Enhance biodiversity
- Provide food and shelter for wildlife
- Structure for dragonflies and midges (fish food)



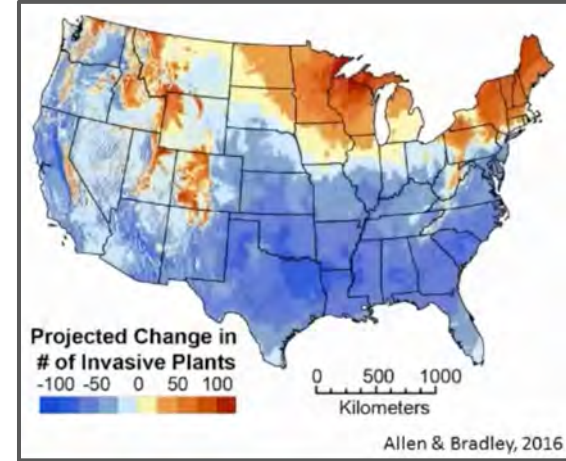
Anthropogenic threats to aquatic plants and those that depend on them



Desire for “clean” swimming areas

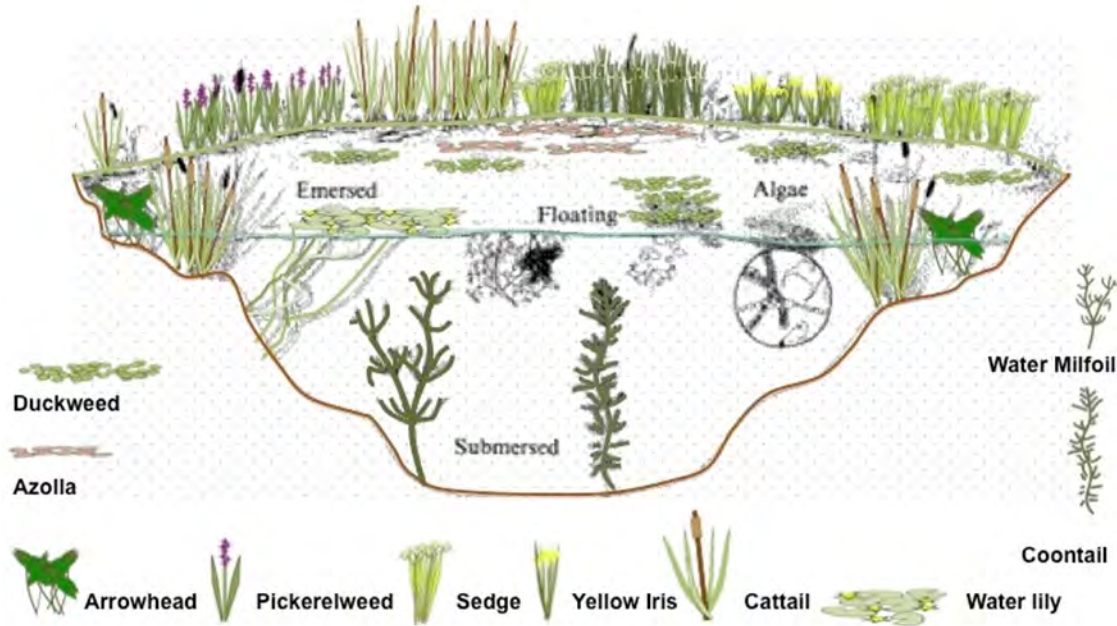


Shoreline development



Climate-driven spread of invasive species

Diverse plant communities are more resilient to environmental stress



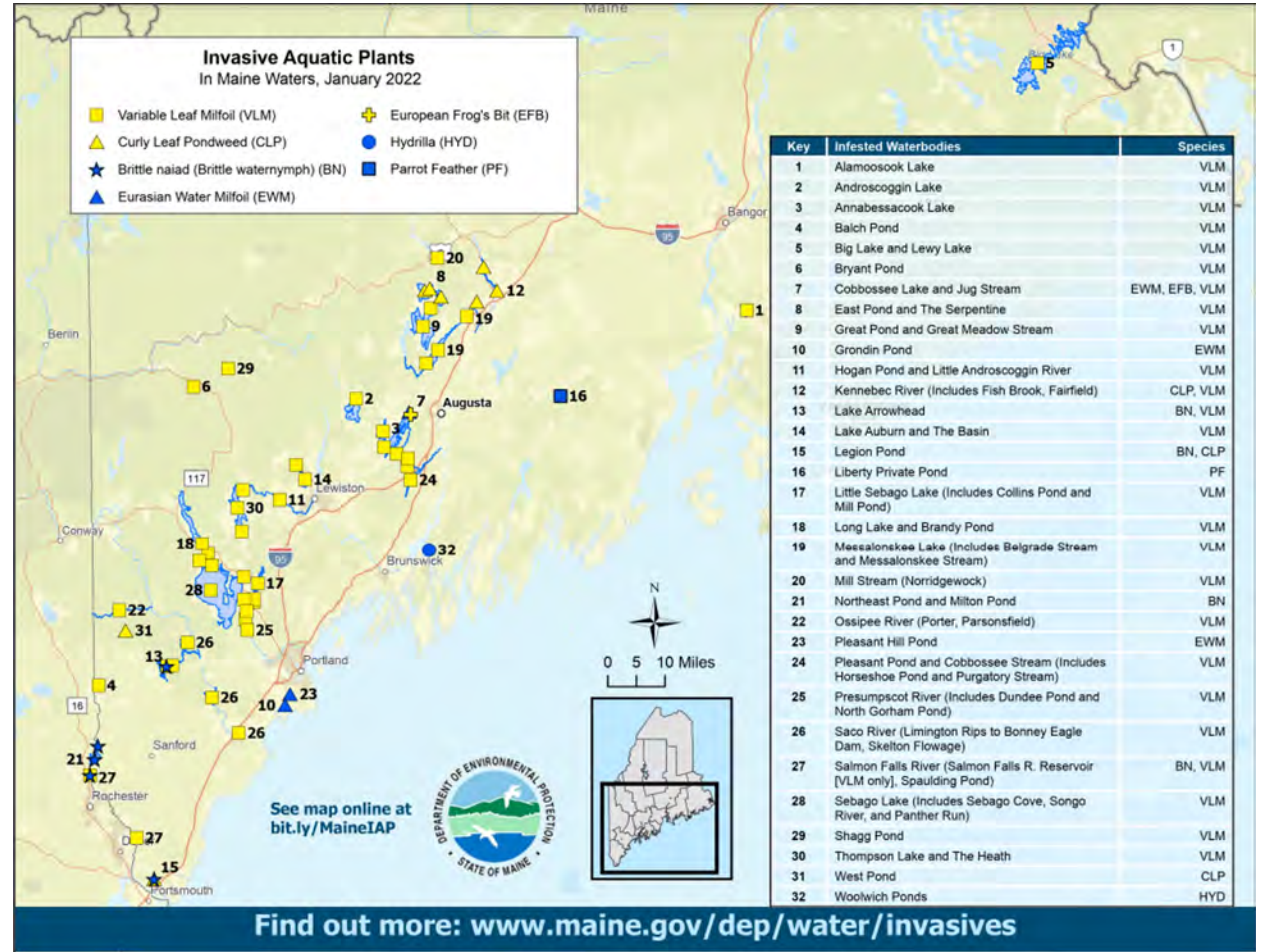
Invasive milfoil in Great Pond



Less than 1% of
Maine's lakes are
infested with one of
"the 11-most unwanted
invasive aquatic plants"

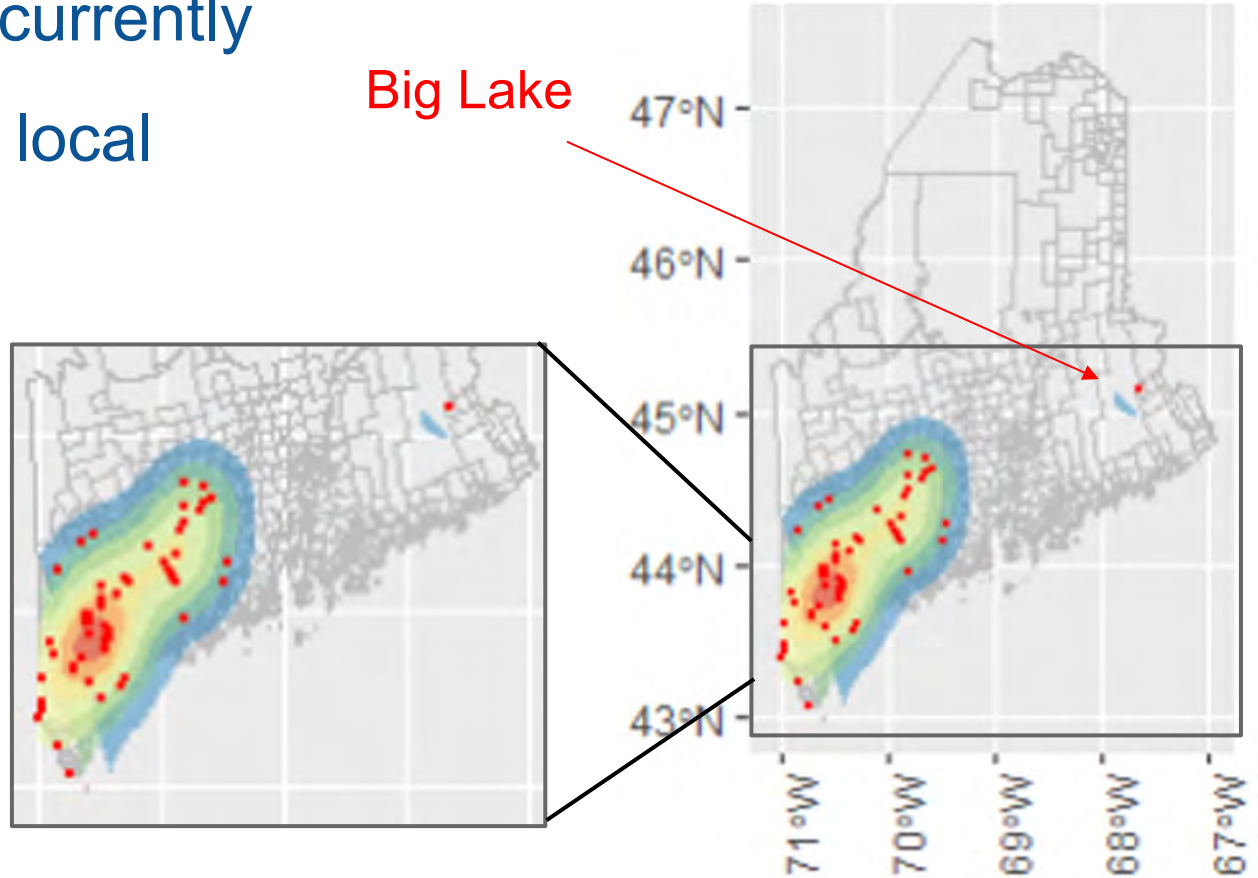
Since 2011, the
number of infestations
has DOUBLED

Most of the infested
water bodies are
popular tourist
destinations and are
close to major
highways



Many lakes are currently
unstewarded by local
groups due to:

- Low population
- Limited funding
- Inaccessibility



Big Lake, Washington County

- Grand Lake Stream area, close to NB, Canada
- Large lake (10,444 AC)
- Part of 17,000 AC system (west branch of Saint Croix River)
- Known worldwide for wilderness beauty & extraordinary fishing.



Early detection is KEY! Need a way to survey aquatic plant communities that is:

- (1) Inexpensive
- (2) Easy
- (3) Informative
- (4) Adaptable



Genetic material obtained directly from environmental samples without any obvious signs of the biological source material (Thomsen and Willerslev, 2015)

Where does eDNA come from?

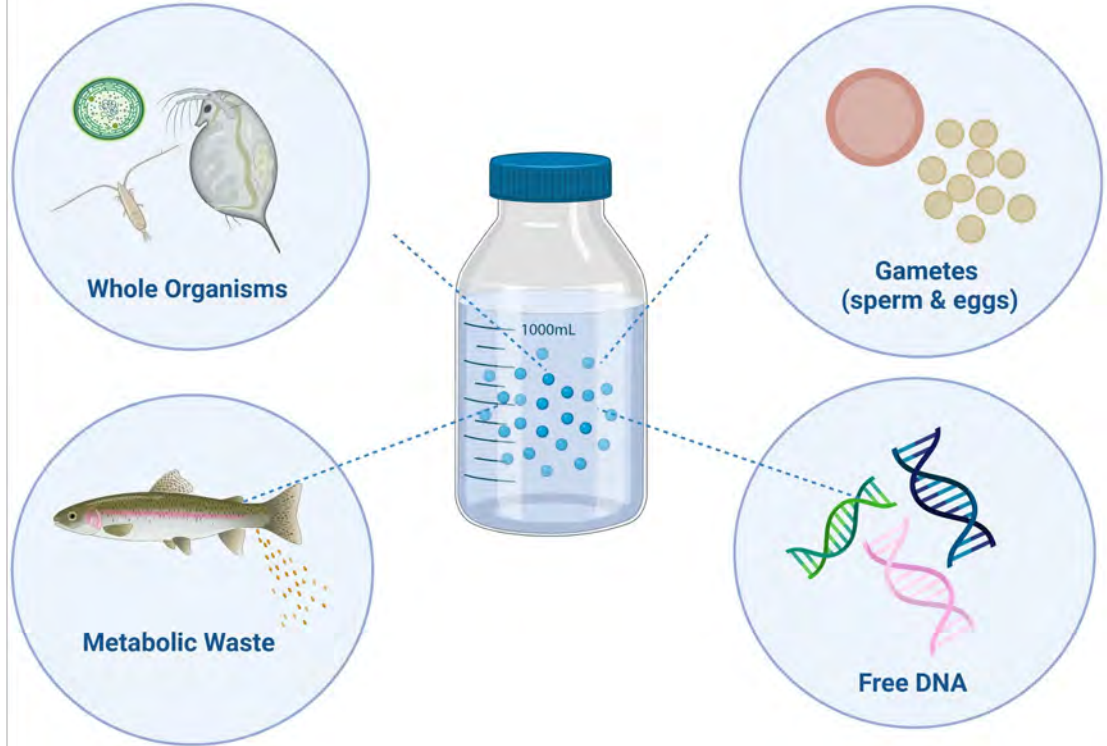
- Cellular decomposition
- Whole shed cells
- Whole microorganisms

Where is eDNA found?

- Water
- Soils
- Air

How is eDNA used?

- Community characterization (metabarcoding i.e. general primers)
- Targeted detection & quantification (qPCR)



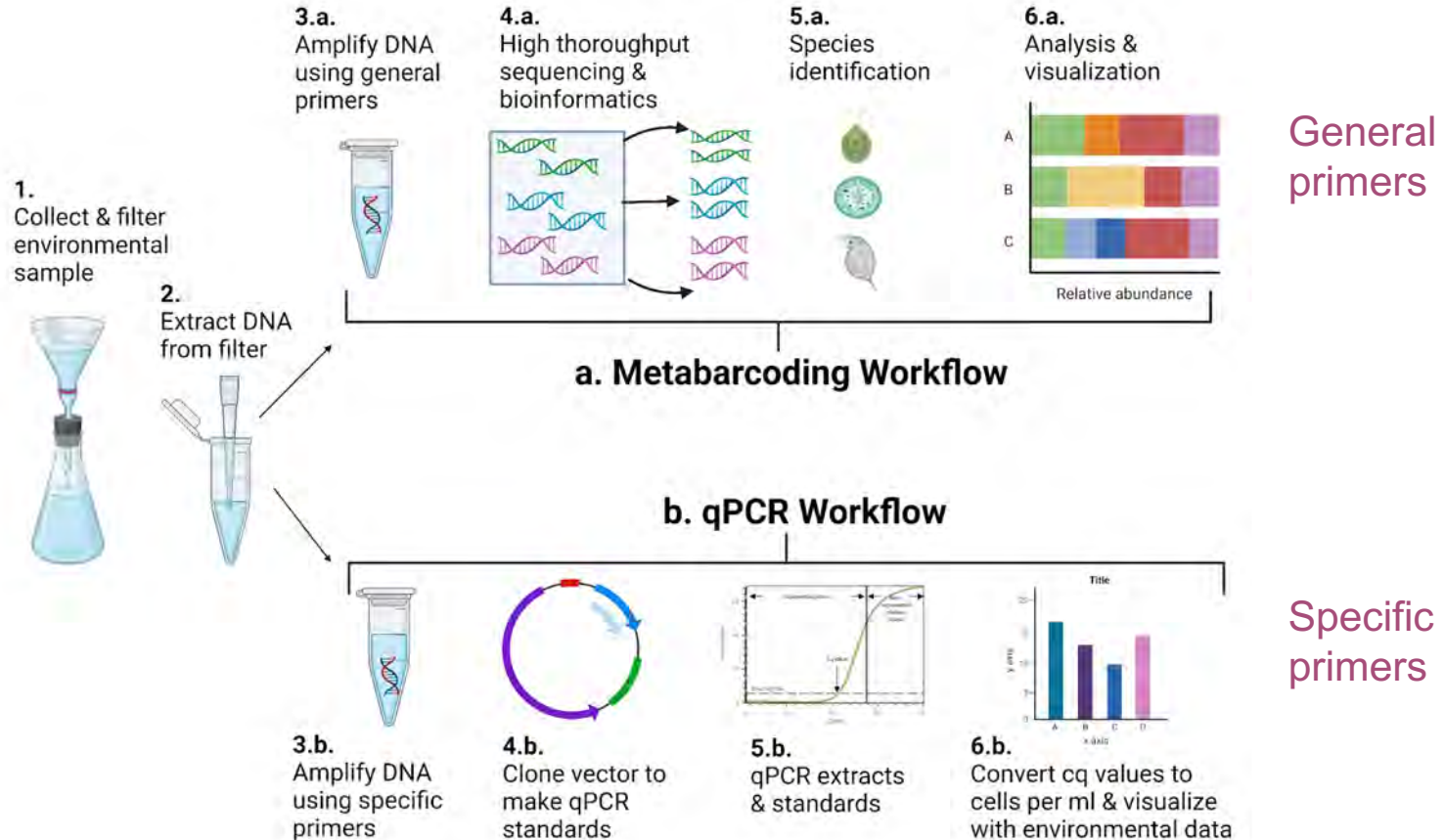
Using a two-tiered approach with eDNA tools

Metabarcoding for whole community composition

qPCR for targeted detection



Using a two-tiered approach with eDNA tools



No PCR = no primers!

Mixed microbial community



DNA
Extraction



Amplicon sequencing



Multiple copies of fragments
from 1 target gene

Metagenomics sequencing



Short sequence
fragments from "all" DNA

1



Develop targeted
qPCR assays for
single-species
detection

2



Build a Maine
metabarcoding
library of native and
invasive aquatic
plants for long-term
monitoring

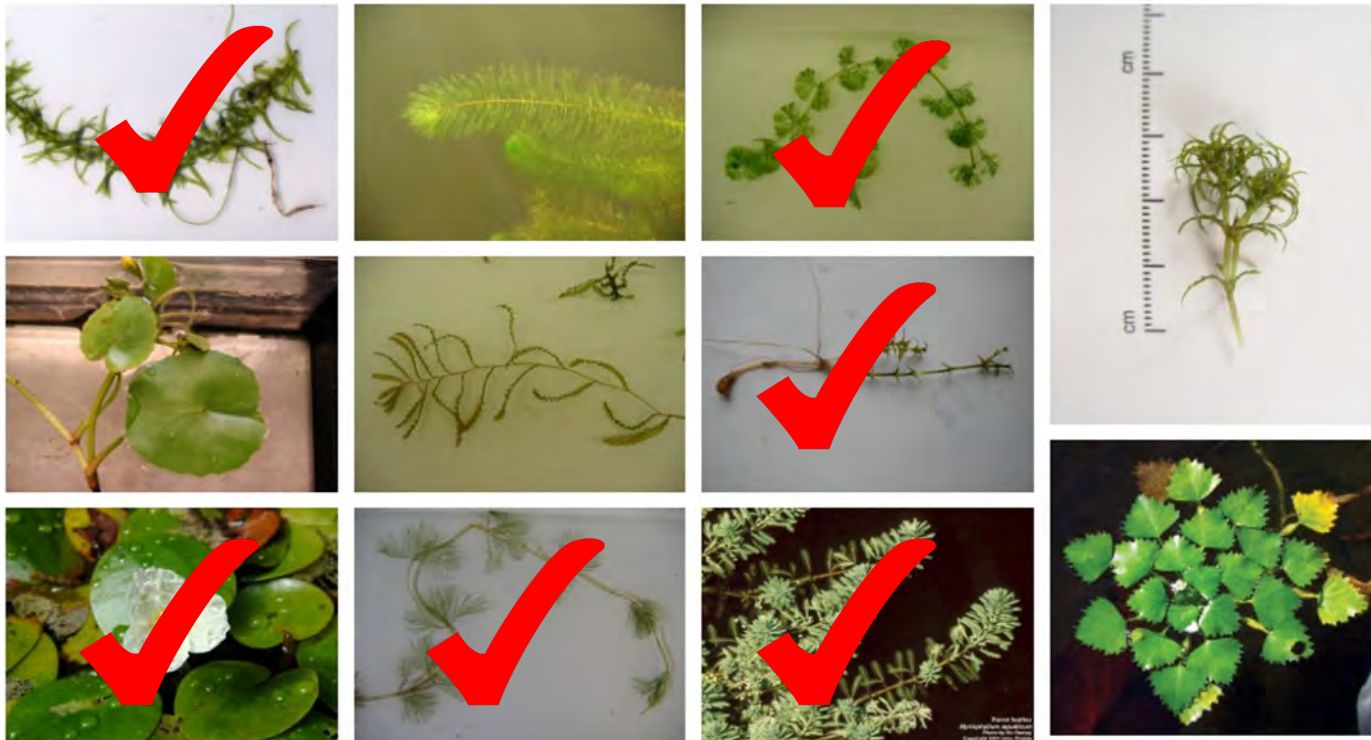
3



Inventory population
diversity statewide



Half of “the 11 most unwanted” have published targeted qPCR primers



Will add
new
invasives
as they
come up





Metabarcoding is all about the library

- Published libraries are not great for aquatic plants
- Aquatic plant sequences from around the globe
- Need to fill our library with our own plant sequences
- Both **native** and **invasive**



Development of an environmental DNA metabarcoding assay for aquatic vascular plant communities

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Abstract

Environmental DNA (eDNA) metabarcodes allow for the simultaneous detection of multiple taxa if the barcode regions meet several key requirements including conserved primer-binding sites, interspecific variability that exceeds intraspecific variability, and relatively short amplicons. Currently, there are no established metabarcoding assays for aquatic vascular plants, which could limit biodiversity assessments and the early detection of alien species. We used a combination of novel and pre-existing primers to generate

New research has shown that aquatic plant eDNA pairs well with traditional surveys

In some cases, eDNA identified more rare species than identified using traditional methods!

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Estimating aquatic plant diversity and distribution in rivers from Jingjinji region, China, using environmental DNA metabarcoding and a traditional survey method

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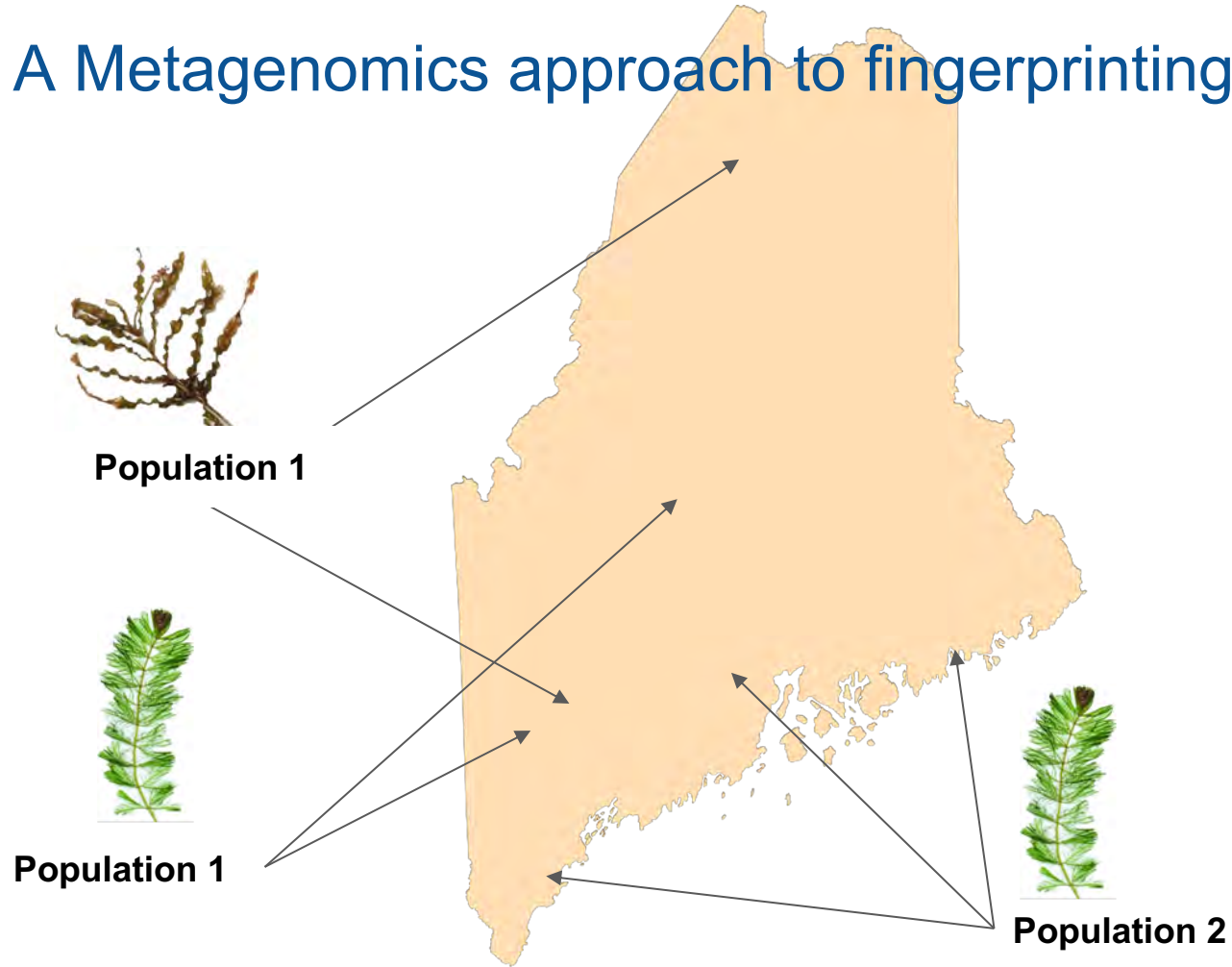
Aquatic plant diversity
Environmental DNA metabarcoding
Traditional survey method

ABSTRACT

Traditional survey methods (TSMs) are difficult to use to perform a census of aquatic plant diversity completely in river ecosystems, and improved aquatic plant community monitoring programs are becoming increasingly crucial with a continuous decline in diversity. Although environmental DNA (eDNA) metabarcoding has been applied successfully to assess aquatic biodiversity, limited work has been reported regarding aquatic plant di-



A Metagenomics approach to fingerprinting



How will the data be used?

- Faster response to new infestations
- Conservation of rare or threatened native species
- To monitor long-term community shifts
- Invasive species monitoring



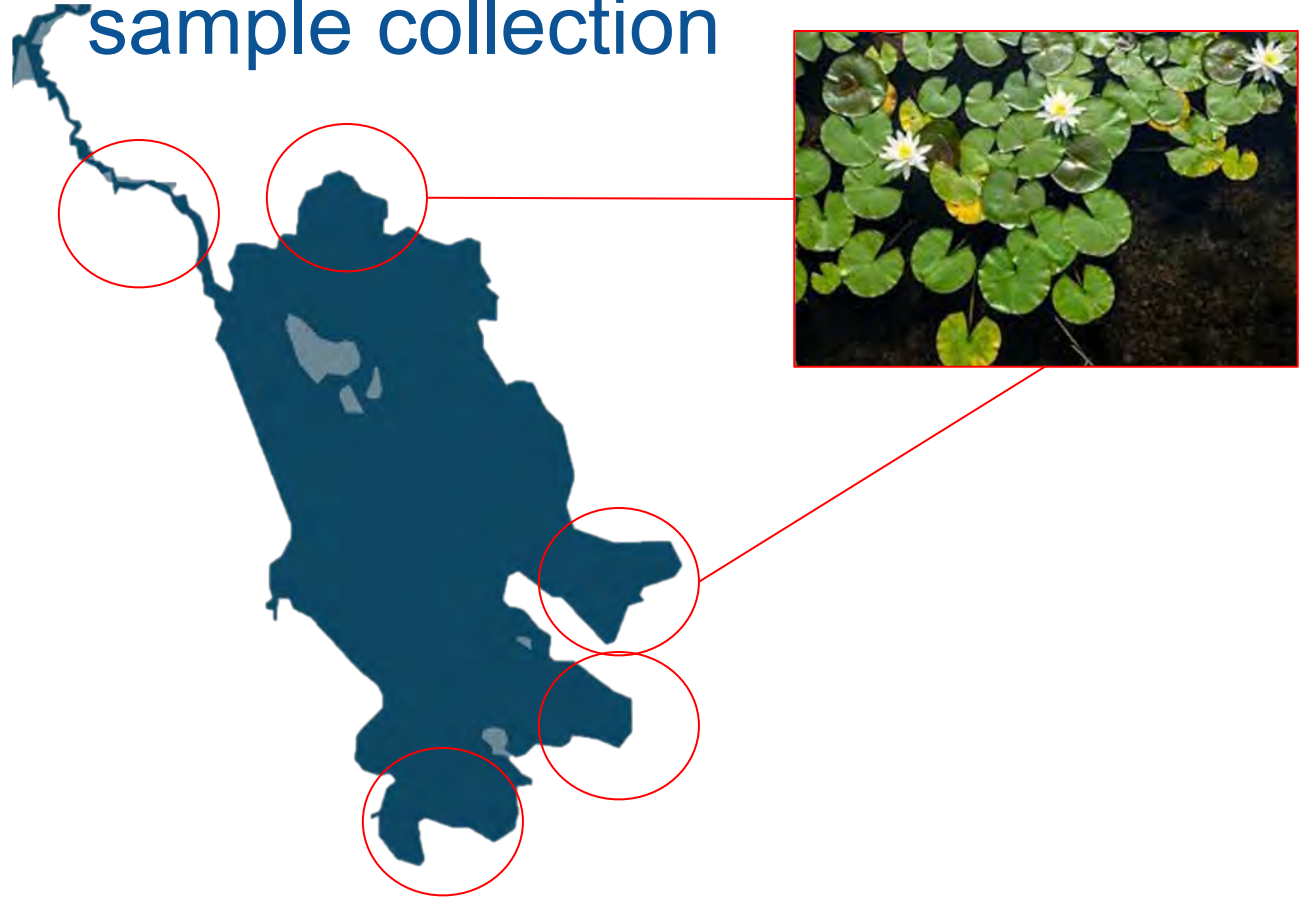
Image credit: Jen Smith Mayo, UMO



Plans for Phase 1

- Design primers for remaining IAPs (Bigelow Labs)
- Test eDNA detection over distance and flow (7 Lakes)
- Start building the library (**open for collaboration!**)

Volunteers will be needed to help with data & sample collection



Step 1.

Identify fertile pockets in the waterbody

Step 2. Inventory all aquatic plants in fertile pockets



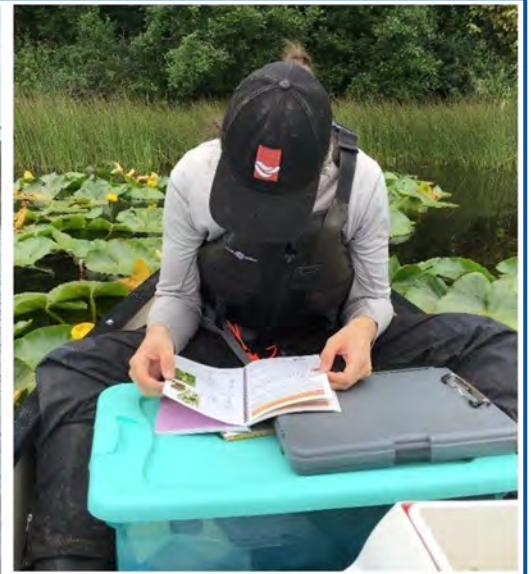
Waterbody _____ Date/s _____ Page _____ of _____

Section 3: Native Plant Inventory For inventory: ✓ = presence; D = dominant; U = uncommon

Native Plant Checklist **	Sector			Common Native Plant Checklist (con't)	Sector		
<i>Typha latifolia</i>				pondweed, small <i>Potamogeton pusilloid</i> spp.			
<i>Sagittaria latifolia</i>				pondweed, spiral-fruited <i>Potamogeton spirillus</i>			
<i>Sagittaria graminea</i>				pondweed, spp. <i>Potamogeton</i> spp.			
<i>Sagittaria</i> spp.				pondweed, variable <i>Potamogeton gramineus</i>			
<i>Utricularia vulgaris</i>				quillwort spp. <i>Isoetes</i> spp.			
<i>Juncus radiata</i>				rush, bayonet <i>Juncus militaris</i>			
<i>Juncus gibba</i>				rush, brown-fruited <i>Juncus pelocarpus</i>			
<i>Najas purpurea</i>				spatterdock <i>Nuphar variegata</i>			
<i>Eleocharis acicularis</i>				spikerush, needle <i>Eleocharis acicularis</i>			
<i>Eleocharis robbinsii</i>				spikerush, Robbin's <i>Eleocharis robbinsii</i>			
<i>Spongia</i> spp.				sponge, freshwater spp.			
<i>Nitella tabernaemontanii</i>				stonewort, spp. <i>Nitella</i> spp.			
<i>Dulichium arundinaceum</i>				three-way sedge <i>Dulichium arundinaceum</i>			
<i>Ranunculus fluitans</i>				water buttercup <i>Ranunculus</i> spp.			
<i>Nymphaea odorata</i>				water lily, fragrant <i>Nymphaea odorata</i>			
<i>Lobelia dortmanna</i>				water lobelia <i>Lobelia dortmanna</i>			
<i>Bidens beckii</i>				water marigold <i>Bidens beckii</i>			
<i>Sium suave</i>				water parsnip <i>Sium suave</i>			
<i>Callitriche</i> spp.				water starwort, spp. <i>Callitriche</i> spp.			
<i>Myriophyllum alterniflorum</i>				water-milfoil, alternate-flowered <i>Myriophyllum alterniflorum</i>			
<i>Myriophyllum tenellum</i>				water-milfoil, dwarf <i>Myriophyllum tenellum</i>			
<i>Myriophyllum farwellii</i>				water-milfoil, Farwell's <i>Myriophyllum farwellii</i>			

Step 3.

- Collect plant samples (alcohol and herbarium presses)
- Collect water for eDNA outside each fertile pocket
- Take pictures
- GPS locate
- Share information





<https://forms.gle/x92PnEgtwYPZWT8E6>

Thank you to all of our partners



Ensuring Our Future



LAKE & WATERSHED
Associates

Extra Slides

Environmental DNA

Manual for Volunteers



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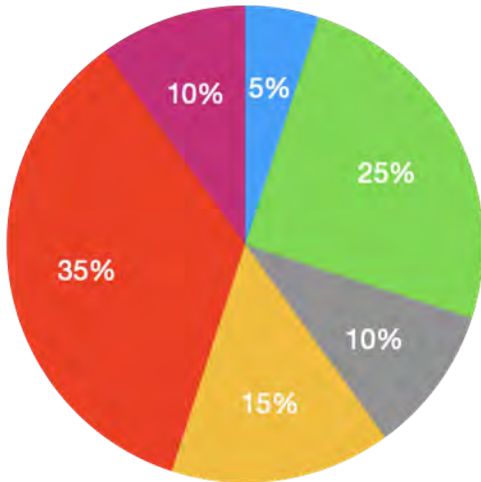
Question:

What can eDNA methods do for Maine Lakes?

Your Answers:

“Invasive Species” and “Help”

What species/communities would you be interested in detecting within Maine lakes? (n=17)



- Don't Know
- Plants
- Invertebrates
- Vertebrates
- Invasive species
- Toxic Species



How Can IPPers Help?



